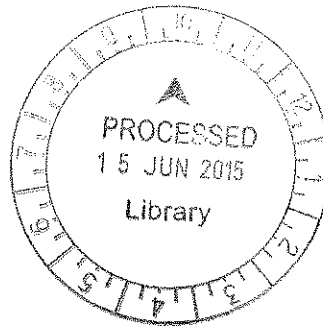


THE EFFECTS OF LACTOSE AND INCUBATION TEMPERATURES ON
SIMILAC 1 AND SIMILAC LF MILK SAMPLES

LIM JIA XIN

DISSERTATION SUBMITTED IN PARTIAL FULFILMENT OF THE
REQUIREMENTS FOR THE DEGREE OF
BACHELOR OF BIOTECHNOLOGY (HONOURS)



FACULTY OF SCIENCE, TECHNOLOGY,
ENGINEERING AND MATHEMATICS
INTI INTERNATIONAL UNIVERSITY
PUTRA NILAI, MALAYSIA

2015

NON-PLAGIARISM DECLARATION

By this letter I declare that I have written this dissertation completely by myself, and that I have used no other sources or resources than the ones mentioned.

I have indicated all quotes and citations that were literally taken from publications, or that were in close accordance with the meaning of those publications, as such. All sources and other resources used are stated in the references.

Moreover I have not handed in a dissertation similar in contents elsewhere.

In case of proof that the dissertation has not been constructed in accordance with this declaration, the Faculty of Science, Technology, Engineering and Mathematics has the right to consider the research dissertation as a deliberate act that has been aimed at making correct judgment of the candidate's expertise, insights and skills impossible.

I acknowledge that the assessor of this item may, for the purpose of assessing this item,

- reproduce this assessment item and provide a copy to another member of the University; and/or,
- communicate a copy of this assessment item to a plagiarism checking service (which may then retain a copy of the assessment item on its database for the purpose of future plagiarism checking).

In case of plagiarism the examiner has the right to fail me and take action as prescribed by the rules regarding Academic Misconduct practiced by INTI International University.

Lim Jia Xin

Name



Signature

111008224

I.D.Number

8/5/2015

Date

ACKNOWLEDGEMENT

I would like to show my appreciation to my dearest supervisor Ms. Lalita Ambigai Sivasamugham for guiding me all along during my final year project. Her motivation, advices and patience have led me to successfully complete the final year project and thesis writing. I would also like to thank her for spending her precious and quality time with me to help me in improvising my project. In addition, I would like express my utmost gratitude to Ms. Emily Quek Ming Poh for her inspirational guidance during the absence of my main supervisor. I would also like to thank Ms. Shiney John for her kind assistance in the statistical analyses based on the data obtained.

Besides, I would also like to thank the laboratory assistants, Ms. Quah Hui Hsien and Mr. Ng Peng Wah for kindly providing advice as well as apparatus and the needed chemicals throughout the project. Without their helping hands and sincere support, I will not be able to complete my project smoothly.

I would also like to take this opportunity to thank my friends, Lam Chi Yan, Yap Ke Xin, Lim Chi Sien, Christopher Leeraj, Kiew Wen Yi, Saktis Paul and Calvin Khoo Chee Feng for giving me the moral support. In addition, I would also like to thank my other course mates who have encouraged me and stood by me at all times. I was able to endure the difficult and stressful periods with their constant support.

Lastly, I would like to thank my parents, my brother and other family members for their love and support. Their care has been a motivation to me to successfully complete this project.

ABSTRACT

Milk spoilage can be determined through chemical and physical analyses. The chemical analysis includes the analysis on the lactic acid content while the physical analysis uses the sensory tests to determine the spoiled milk. Most spoilage microbes such as *Bacillus*, *Proteus*, *Staphylococcus* and *Lactobacillus* grow easily in milk using the available nutrients such as lactose to produce lactic acid as the main end-product. Thus, the aim of this research was to study the effects of the lactose in the lactose-containing milk (Similac 1) and lactose-free milk (Similac LF) and the incubation temperature of 4°C, 25°C, 37°C and 60°C on the lactic acid content (g/100 mL) and the CFU/mL. The incubated milk was also analyzed by observing the curd formation, mold growth and the colour change of whey. The lactic acid content of the incubated milk was determined using the titration test with 0.1M NaOH. The CFU/mL was determined using the pour plate procedure and lastly, gram staining was done to identify the gram reaction of the spoilage microbes isolated from the pour plate method. The Analysis of Variance (ANOVA) indicated that the lactose in the milk and the incubation temperatures affected the lactic acid concentration as well as the CFU/mL of the incubated milk samples. In addition, the amount of the lactic acid also affected the CFU/mL of the incubated milk samples as the acidic condition probably inhibited the growth of acid-labile bacterial cells. Gram staining reactions revealed more gram-positive bacteria in the milk samples compared to gram-negative bacteria. The preliminary chi-square analysis of the physical observation indicated that only the incubation temperature affected some of the physical characteristics of the milk

TABLE OF CONTENT

| | PAGE |
|--|-------------|
| DECLARATION | ii |
| ACKNOWLEDGEMENT | iii |
| ABSTRACT | iv |
| TABLE OF CONTENT | v |
| LIST OF TABLES | viii |
| LIST OF FIGURES | ix |
| LIST OF ABBREVIATIONS | x |
| CHAPTER | |
| 1. INTRODUCTION | 1 |
| 2. LITERATURE REVIEW | 3 |
| 2.1 Milk | 3 |
| 2.2 Milk Spoilage | 4 |
| 2.2.1 The Four Stages of Milk Spoilage | 4 |
| 2.2.2 Chemical and Physical Changes of Spoiled Milk | 5 |
| 2.3 Factors That Lead to Spoilage of Milk | 5 |
| 2.3.1 Physical-based: Temperature | 6 |
| 2.3.2 Chemical-based | 6 |
| 2.3.2.1 The Carbohydrates | 6 |
| 2.3.2.2 The Milk Fat | 7 |
| 2.3.2.3 The Milk Protein | 8 |
| 2.3.3 Bacteriological-based: Contamination of Raw Milk | 8 |
| 2.4 Methods to Indicate Milk Spoilage | 9 |
| 2.4.1 Methylene Blue Reduction Test | 9 |
| 2.4.2 pH Measurements | 10 |
| 2.4.3 Cell Count | 10 |
| 2.4.4 Observation of Milk Spoilage through Physical Appearance | 11 |
| 2.5 Identifying The Spoilage Bacteria Strain In Spoiled Milk Through Gram Staining | 12 |
| 2.6 Methods to Avoid Milk Spoilage During Packaging Process | 12 |
| 2.6.1 Pasteurization | 12 |
| 2.6.2 Ultra-high Temperature (UHT) Treatment | 13 |
| 3. MATERIALS AND METHODS | 14 |
| 3.1 Materials | 14 |
| 3.1.1. Source of Apparatus and Materials | 14 |
| 3.1.2 Condition of Apparatus and Materials Used | 14 |

| | | |
|-----------|---|-----------|
| 3.2 | The Milk Samples | 14 |
| 3.2.1 | Milk Samples Preparation and Incubation | 14 |
| 3.3 | Detection of Milk Spoilage | 16 |
| 3.3.1 | pH Value | 16 |
| 3.3.2 | Determination of Lactic Acid Content | 17 |
| 3.3.3 | Plate Count | 17 |
| 3.3.4 | The Physical Observation of Incubated Milk | 19 |
| 3.4 | Gram Staining – A Preliminary Identification of The Spoilage Microbes | 19 |
| 4. | RESULTS AND ANALYSIS | 21 |
| 4.1 | Lactic Acid Content in Milk | 21 |
| 4.1.1 | The Lactic Acid Content (g/100 mL) in the incubated Lactose-Free (LF) and Lactose-Containing (LC) Milk at Each Incubated Temperature | 21 |
| 4.1.2 | The Overall Comparison of Lactic Acid Content of Lactose-Containing (LC) Milk and Lactose-Free (LF) Milk at Different Incubation Temperatures | 24 |
| 4.1.3 | The Two-Way ANOVA on The Lactic Acid Content | 26 |
| 4.2 | Plate Count | 27 |
| 4.2.1 | Analysis of CFU/mL of The Lactose-Containing (LC) and Lactose-Free (LF) Milks at Each Incubation Temperature | 27 |
| 4.2.2 | The Overall Comparison of CFU/mL Analysis of The Incubated Lactose-Containing (LC) Milk and Lactose-Free (LF) Milk | 29 |
| 4.2.3 | The Two-Way ANOVA Test on CFU/mL | 30 |
| 4.3 | Physical Observation | 31 |
| 4.3.1 | Curd Formation | 32 |
| 4.3.2 | Colour Changes of Whey | 33 |
| 4.3.3 | Mold Growth | 34 |
| 4.4 | Gram Stain | 35 |
| 5. | DISCUSSION | 39 |
| 5.1 | Lactic Acid Concentration (g/100 mL) | 39 |
| 5.2 | Cell Growth in Milk | 41 |
| 5.3 | The Spoilage Bacteria | 43 |
| 5.4 | Physical Observation | 44 |
| 6. | CONCLUSION AND RECOMMENDATIONS | 47 |
| | REFERENCES | 49 |
| | APPENDICES | 56 |
| | Appendix I | 56 |
| | Appendix II | 58 |
| | Appendix III | 60 |
| | Appendix IV | 63 |
| | Appendix V | 65 |
| | Appendix VI | 70 |

Appendix VII
Appendix VIII

74
76

LIST OF TABLES

| Table | | Page |
|-------|--|------|
| 1 | Storage of Milk | 16 |
| 2 | The ANOVA of the effects of the lactose content (milk types) and incubation temperatures on lactic acid content of milk. | 26 |
| 3 | The ANOVA of the effects of the lactose content (milk types) and the incubation temperatures on the CFU/mL. | 31 |
| 4 | The chi-square test of independence or association between the lactose content (type of milk) and the presence of curd. | 32 |
| 5 | Chi-square test of independence or association between the incubation temperature of milk and the presence of curd. | 33 |
| 6 | Chi-square test of independence or association between the lactose content (milk types) and the colour changes of whey. | 33 |
| 7 | Chi-square test of independence or association between the incubation temperature of milk and the presence of curd. | 34 |
| 8 | Chi-square test of independence or association between the lactose content (milk types) and mold growth. | 35 |
| 9 | Chi-square test of independence or association between the incubation temperature and the mold growth. | 35 |
| 10 | The gram reaction and the bacteria morphology of the incubated LF and LC milk. | 38 |

LIST OF FIGURES

| Figure | | Page |
|--------|--|------|
| 1 | The content of lactic acid of LC and LF milks incubated at 4°C. | 22 |
| 2 | The content of lactic acid of LC and LF milks incubated at 24°C. | 23 |
| 3 | The content of lactic acid of LC and LF milks incubated at 37°C. | 23 |
| 4 | The content of lactic acid of LC and LF milks incubated at 60°C. | 24 |
| 5 | The content of lactic acid of LC milks incubated at four different temperatures; 4°C, 25°C, 37°C and 60°C. | 25 |
| 6 | The content of LF milks incubated at four different temperatures; 4°C, 25°C, 37°C and 60°C. | 25 |
| 7 | The CFU/mL LC and LF milks incubated at 4°C. | 27 |
| 8 | The CFU/mL LC and LF milks incubated at 25°C. | 28 |
| 9 | The CFU/mL LC and LF milks incubated at 37°C. | 28 |
| 10 | The CFU/mL LC and LF milks incubated at 60°C. | 29 |
| 11 | The CFU/mL of LC milks incubated at 4°C, 25°C, 37°C and 60°C. | 30 |
| 12 | The CFU/mL of LF milks incubated at 4°C, 25°C, 37°C and 60°C. | 30 |
| 13 | Gram positive bacilli found in the LC milk incubated at 25°C. | 36 |
| 14 | Gram positive bacilli found in the LF milk incubated at 25°C. | 36 |
| 15 | Gram positive bacilli found in the LF milk incubated at 37°C. | 37 |
| 16 | Gram positive and gram negative bacilli bacteria found in the LC milk incubated at 37°C. | 37 |
| 17 | Gram positive cocci bacteria found in the LC milk incubated at 60°C. | 38 |

LIST OF ABBREVIATIONS

| | |
|--------------------|------------------------------------|
| α | Significance level |
| ANOVA | Analysis of variance |
| $^{\circ}\text{C}$ | Degrees Celsius |
| CFU/g | Colony forming unit per gram |
| CFU/mL | Colony forming unit per milliliter |
| CO_2 | Carbon dioxide gas |
| <i>df</i> | Degree of freedom |
| <i>E. coli</i> | <i>Escherichia coli</i> |
| $^{\circ}\text{F}$ | Fahrenheit |
| FFA | Free fatty acid |
| G | Gram |
| g/100 mL | Gram per hundred milliliter |
| GLM | General linear model |
| H_0 | Null hypothesis |
| H_1 | Alternative hypothesis |
| H_2 | Hydrogen |
| LC | Lactose-containing |
| LF | Lactose-free |
| M | Mole |
| mL | Milliliter |
| <i>MS</i> | Mean square |
| μL | Microliter |
| <i>n</i> | Number of replicates |
| NaOH | Sodium hydroxide |

| | |
|-----|------------------------|
| SS | Sum of squares |
| UHT | Ultra-high temperature |
| UP | Ultrapasteurization |
| V/V | Volume to volume |

CHAPTER 1

INTRODUCTION

Milk is a nutritious food (Singh, Kaushal, Tyagi & Sharma, 2011) that is important for all including expectant mothers and growing children (Srujana, Rajender, Krishna & Ram, 2011). Milk contains fats, proteins, carbohydrates, vitamins and minerals and high water content (Singh et al., 2011; Al-Shammary, 2013). This makes milk a suitable growth medium for microorganisms (Al-Shammary, 2013). There are two main groups of microorganisms found in milk; the spoilage and the pathogenic microorganisms. The spoilage microorganisms utilize the milk components such as lactose to synthesize compounds such as, lactic acid (Singh et al., 2011; Al-Shammary, 2013). Meanwhile, the pathogens will produce toxins or harmful substances leading to milk-borne diseases such as typhoid fever (Singh et al., 2011; Al-Shammary, 2013). In some cases, bacteria such as *Bacillus cereus* can be both pathogenic and spoilage microbes (Al-Shammary, 2013).

There are various factors that cause spoilage of milk. One of the main factors is the high moisture level and the availability of dissolved oxygen that enables the growth of both aerobic and facultative anaerobic microorganisms (Lu et al., 2013). Besides that, improper storage temperature of milk will also cause the milk to spoil faster. Studies show that temperatures between 20-30°C to increase the metabolic rate of the microbes causing them to grow faster in milk (Ranken, Christopher & Kill, 1997; Samarzija, Zamberlin & Pogacic 2012). Besides that, unsterilized milking machines and non-pasteurized milk will also cause the milk to spoil faster (Samarzija et al., 2012).

Microbial growth and the metabolites produced changes the appearance, taste and odor of milk. The spoiled milk can turn bitter, rancid, sour and malty in taste (Duyvesteyn, Shimoni & Labuza, 2011; Al-Shammary, 2013). It is hard to measure milk spoilage accurately (Lu et al., 2013). However, there are certain methods that can be used to detect milk spoilage with higher accuracy such as pH testing, methylene blue reduction with amperometric sensor, megnetoelastic, gas-sensor array,

infrared spectroscopy and protein or fat count (Lu et al., 2013; Al-Qadiri, Al-Holy, Cavinato & Rasco, 2007). The infrared spectroscopy method is the most accurate and it has broad spectrum of detection with high sensitivity. The results of the spoiled milk can be obtained within 4 minutes. However, the cost for buying this infrared spectroscopy is high and the running cost for the infrared test is low (Lu et al., 2013).

To date, there isn't a concrete research to study the relation of the lactose content and the incubation temperatures on the incubated milk. Hence, this study was aimed to provide a better insight on the effects of the lactose containing milk and the lactose free milk to the milk by looking at the lactic acid content of the milk samples incubated at 4 temperatures, 4°C, 25°C, 37°C and 60°C. The effects were determined using titration (determining the lactic acid content), plate counting and physical observation based on the colour of the whey, the curd formation and mold growth. Gram-staining was done on the bacterial colonies to identify the gram reaction of the spoilage bacteria found in the milk.

There were two main hypotheses in this study which were:

H₀: There were no significant effects of the lactose (the type of milk) and the incubation temperatures on the lactic acid content (g/100 mL) and CFU/mL

H₁: There were significant effects of the lactose (the type of milk) and the incubation temperatures on the lactic acid content (g/100 mL) and CFU/mL

CHAPTER 2

LITERATURE REVIEW

2.1. MILK

Milk is produced in the mammary glands of mammals (Codex, 2011). The colour of milk varies from white to yellow. The difference in the colour is determined by the content of carotene in milk fat (Bylund, 1995). Milk is a very nutritious as it consists of nutrients which are needed by our body such as high protein content, riboflavin, magnesium, vitamin B12, selenium, pantothenic acid and calcium (FAO, 2013). Milk also contains 87.3 - 88.1 % (V/V) of water (FAO, 2013). It is either directly consumed or processed before consumption, or processed into different products (Codex, 2011).

Milk can be categorized into several classes. It can either be classified according to processing procedures, for examples the ultra-high-temperature treated milk (UHT-milk), pasteurized milk or sterilized milk. It can also be classified by its fat content such as low-fat milk, full-cream milk or skimmed milk. In addition, milk can be classified according to the sugar content for instance lactose-containing milk (the usual milk), and the lactose-free milk. Lactose content is generally around 4.8 – 5.2% in milk. In fact, lactose is the major carbohydrate found in milk (Choi, Lee & Won, 2007).

Lactose-free milk is produced for the consumption of lactose intolerant individuals. This is because these individuals are unable to digest lactose as they produce insufficient lactase, the enzyme responsible for digesting lactose (Dairy Council of California, 2015). Lactose-free milk is usually produced by the process of enzyme hydrolysis. In this process, the milk is treated by microbial β -galactosidase so that the lactose content in the milk can be broken down into glucose and galactose (FDA, 2011). The mixture of glucose and galactose causes the lactose-free milk to taste sweeter compared to the normal lactose-containing milk (Choi, Lee & Won, 2007). Due to the sweet taste, Tossavainen and Sahlstein (2003) used ultrafiltration

technique to remove permeate, which consists of milk lactose and milk salt, from the milk. The milk salt in the permeate was recovered again through reverse osmosis and nanofiltration and added back into the reconstituted milk which is the milk without permeate. The reconstituted milk with low lactose content is then treated with β -galactosidase in order to produce less-sweet lactose-free milk (Tossavainen and Sahlstein, 2003).

2.2. MILK SPOILAGE

Milk spoilage refers to the changes or deterioration of the physical appearance, volatility, taste and odor (Kim, Hardy, Novak, Ramet & Weber, 1983; Al-Shammary, 2013) of the milk. These changes are caused by the growth of microbes in the milk (Kim et al., 1983). Some of the common pathogens found in milk samples include species of *Bacillus*, *Proteus*, *Staphylococcus*, *Salmonella* as well as *E. coli* (Singh et al., 2011). Both the nutritional composition of the milk as well as the storage temperature encourage these microbes to grow and spoil the milk.

2.2.1. The Four Stages of Milk Spoilage

There are four stages of milk decay which are rancid, curdling, coagulation and dry (F&N Magnolia, 2013). Rancidity refers to the slight souring of milk. Curdling refers to the formation of curd (F&N Magnolia, 2013; Antunes et al., 2014) that is caused by the renin-like enzyme secreted by certain thermophiles such as *Proteus* and *Bacillus* (Jay, Loessner & Golden, 2005; Al-Shammary, 2013). In both the rancid and curdling phases, the milk is still consumable but it is not advisable (F&N Magnolia, 2013). However, the milk is no longer consumable if it is coagulated as mold is usually noted to grow at this phase. The milk can become dehydrated and turns chalky and hard when it comes to the dry phase due to the vaporization of water in the milk.

There are also other characteristics in milk that indicates milk spoilage, for example, gas production in milk. Production of gas such as carbon dioxide (CO₂) and hydrogen (H₂) by the lactic acid bacteria will cause clot formation and foaming (Al-Shammary, 2013). The appearance of ropy texture (viscous-slimy texture) also indicates spoilage of milk. The formation of ropy texture is caused by the production